DEVELOPMENT OF A 3-D-PRINTED MOUSE PHANTOM TO REPLACE CURRENT MOUSE ANIMAL MODEL

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Scientific paper https://doi.org/10.2298/NTRP2402121K

Evaluating the radiation dose of target organs of a laboratory mouse requires a glass dosimeter to be surgically inserted at the irradiated location. However, precisely inserting the glass do simeter at the same location in different mice is rarely achieved, reducing the reliability of the measured radiation dose. To address this limitation, 3-D mouse phantom was developed using computed tomography scanning and 3-D printing technology. The radiation dose of target organs was assessed using four mouse models: laboratory mouse, 3-D mouse phantom, Monte Carlo N-Particle (MCNP) 3-D phantom, and MCNP simulation. In all the experiments, the brain, heart, lungs, and abdomen were irradiated with 100 mGy of measured air kerma at a 6 mGyh⁻¹ air kerma rate. A small volume glass dosimeter was inserted into the mouse models to assess the radiation dose, and the reliability of the glass dosimeter reading system was evaluated using the dose-response curves. The dose values of the laboratory mouse and 3-D-printed mouse phantom were found to differ by up to 3.3 %. This study provides a method to accurately measure the radiation dose to target organs, enhancing the reliability of pre-experiments. the measured radiation dose. To address this limitation, 3-D mouse phantom was developed
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target organs was assessed using four mouse mo

Owing to the recent increase in the utilization of 3-D printers, research on the use of output materials and products is being increasingly conducted in the medical field. Although many studies employ 3-D- printed mice, no research has been found on radiation dose evaluation of target organs in mice with a non-breakable traceability ret rospective assessment chain. In this study, a realistic 3-D mouse phantom was created using computed tomography (CT) scanning and 3-D printing techniques [1-3]. The CT was used to capture the internal organs of the laboratory mouse to produce the $3-D$ phantom $[4-7]$. The 3-D mouse phantom was 3-D printed using acrylonitrile butadiene styrene (ABS), a material that is actively used as a tissue equivalent. The radiation dose of target organs was assessed using four mouse models: laboratory mouse, 3-D mouse phantom, Monte Carlo N-particle (MCNP) 3-D phantom, and MCNP simulation. Experiments were performed at the Dongnam Institute of Radiological and Medical Sciences (DIRAMS), an internationally accredited calibration and testing laboratory that belongs to the Korea Laboratory Accreditation Scheme (KOLAS). The absorbed radiation dose of

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INTRODUCTION the four mouse models was evaluated using the traceability of the calibrated ion chamber at the Korea Research Institute of Standards and Science (KRISS). A total of 100 mGy was irradiated using a $137Cs$ gamma radiation field at a biologically low dose rate of 6 mGyh⁻¹ to evaluate the dose irradiated inside the mouse models with a radio photo-luminescent (RPL) glass dosimeter and reading system $[8-10]$. The radiation dose in target organs was assessed and compared in the 3-D mouse phantom, MCNP 3-D phantom, MCNP simulation, and laboratory mouse.

MATERIALS AND METHODS

This study conducted dose assessments of internal organs on four mouse models: laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simulation. The target organs for dose evaluation were the brain, heart, lungs (left and right), and abdomen, as well as organs that can confirm the effect of radiation attenuation in tissues. The laboratory mouse was dissected, and glass dosimeters were inserted into each target organ locations for irradiated dose assessments by gamma radiation which was emitted at 662 keV from ¹³⁷Cs source of each target organ.

To produce the $3-D$ mouse phantom, a 25.15 g laboratory mouse was CT scanned (Light SpeedTMRT16, GE Healthcare) in the T-B direction at 1.25 mm, P-A, and L-R direction at 2.5 mm intervals of a 512 matrix. Using the MIMICS software, a software used for 3-D printing, the captured DICOM images were bound by contour differences in the total body, brain, heart, lungs (left and right), and abdomen and were transformed into an STL output file statement $[11]$. A CT reader expert manually distinguished each mouse organ from the DICOM with a density of 1.05 gcm⁻³, a tensile strength of 50 MPa, file. Each organ was made with a cylinder-shaped hole $(0.3 \text{ mm in diameter and } 9.7 \text{ mm long})$ to insert a glass dosimeter (GD-301, Asahi Techno Glass) for dose assessment in the STL file, as shown in fig. 1. For the lung, a 0.3 gcm⁻³ volume was applied by physically creating

an internal space for density in the 3-D printing process. When a glass dosimeter was inserted, its volume occupied the entire internal lung space, therefore, the effect of lung density was negligible in assessing the radiation dose. The 3-D printed mouse phantom was assembled for glass dosimeter insertion, and a cross section was made at a perpendicular in the beam direction from the centre of glass dosimeter position.

The 3-D phantom was made using ABS material and a multi-body output via the poly-jet method [8]. For comparison, a linear algebra-based mouse was simulated using MCNP 6.2, and a MCNP 3-D phantom was produced using a 3-D printer as shown in fig. 2 [12]. The purpose of dose assessment by a linear algebra-based mouse

Figure 1. The CT scan of a laboratory mouse (a), STL file for 3-D printing (b), and $3-D$ mouse phantom (c)

Figure 2. Geometric shape of linear algebra- based MCNP simulation (a) and MCNP 3-D phantom (b)

 (b)

phantom is to secure the reliability of the computational simulation data by comparing the measurement results
and simulation results. A MCNP 3-D phantom of linear algebra is a phantom that can represent mice of different
size. The MCNP 3-D phantom was also prefabricated,
likewi and simulation results. A MCNP 3-D phantom of linear algebra is a phantom that can represent mice of different size. The MCNP 3-D phantom was also prefabricated, $\frac{1}{8}$ ¹⁵ likewise the 3-D mouse phantom.

A hole was built inside the $3-D$ mouse phantom $\frac{1}{10}$ to evaluate the radiation dose, such that two glass dosimeters on both sides of the spine could be mounted $\left\| \right\|$ based on CT images. The small cylindrical glass dosimeter inserted into the phantom was a silvery active phosphate comprising P (31.55 %), O (51.16 %), Al $\frac{1}{0}$ (6.12%) , Na (11.0%) , and Ag (0.17%) . It was 1.5 mm wide and 8.5 mm long. Glass do sime ters with response stability, compact size, and readability are widely used in field of gamma dose assessment as passive detectors. The glass dosimeter, which was irradiated, underwent 30 minutes of pre-processing at 70° C. The radiation dose was evaluated using a reader (FGD-1000SE, Asahi Techno Glass) with a holder size 2.8 mm wide and 9.5 mm long. The glass dosimetry system was calibrated by producing a dose-response curve using a glass dosimeter that maintains traceability that was standard gamma irradiation at 6 mGy and 2 Gy by ¹³⁷Cs source at KRISS.

It is important not only to accurately read the glass do simetry system, but also to preserve the reliability of the measurement of the gamma dose rate to irradiate the phantom during the experiment. To retain the reliability of the irradiation dose rate, a radiation detection system that maintains traceability was used and compared with the results of MCNP simulation.

A standard calibrated EXRADIN ion chamber (A3, Standard Imaging) with traceability from the KRISS was used to evaluate the air kerma rate from the 137Cs irradiator source (370 GBq, Chiyoda Technol. Corp.) to the detection positions at $1-5$ m. Low-dose irradiation was performed with a total irradiation dose of 100 mGy at a rate of 6 mGyh⁻¹ at 2 m from the $137Cs$ source $[8, 9]$. Low-dose irradiation of the 3-D mouse

Figure 3. Irradiation dose assessment using an ion chamber with traceability and MCNP simulation

phantom was also conducted after dose evaluation of the photon irradiation devices using a standard calibrated A3 ion chamber [13-15]. The dose rate, according to the distance from the irradiator to the detection position, was evaluated by experimental results and MCNP simulation, as shown in fig. 3 [8, 9].

Figure 4 illustrates the experimental set-up for the irradiation experiments. The radiation direction of the laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simulation samples was P-A with positioning on a circular arc-shaped mouseapartment system. The system built in the DIRAMS is designed to irradiate an uniform dose for each position. The distance from the source to the heart of the mouse models was 2 m. The MNCP simulation was set up for each irradiation experiment in ABS plastic cases that had a length, width, height, and thickness of 13, 12, 14, and 0.3 cm, respectively $[16]$.

The laboratory mouse was dissected under the same conditions as the 3-D mouse phantom and MCNP 3-D phantom, and glass dosimeters were inserted into the target organs. After completion of gamma irradiation of a total of 100 mGy , the dose of the glass dosimeter was evaluated by own glass do simetry system.

Figure 4. Experimental set-up for irradiation of laboratory mouse (a), MCNP 3-D phantom (b), and 3-D mouse phantom (c)

In this study, a 3-D mouse phantom was created printing technology to evaluate radiation dose measurements of target organs. The radiation dose and uncertainty of the $137Cs$ radiation units, measured at a distance of 2 m from the source and radiation point, were 6.49 mGyh⁻¹ and 3.1 % (approximately 95 % confidence levels and $k = 2$), respectively, and the total radiation dose at the radiation point was $100 \,\mathrm{mGy}$ [13, 14]. The extended uncertainties of the irradiation dose included measurement uncertainty of the ion chamber, electrometer, thermometer, barometer, beam uniformity and reproducibility of detector location. Table 1 shows the dose reading results of the laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simulation after the completion of the 100 mGy irradiation.

Because there was measurement uncertainty in all the experiments, the measurement uncertainty was evaluated more accurately in the glass dosimetry system using a small-volume element. In addition to the uncertainty of the $137Cs$ radiation units, the elements of relative measurement uncertainty, based on the uncertainties of the dose measurement detector system, included uncertainty of the ion-chamber, ionization ammeter, barometer, thermometer, beam homogeneity, ion re combination, interpolation function, and position repeatability. Additionally, the uncertainty of the glass do simetry system included the standard irradiation of glass dosimeters, reading repeatability, and the interpolation function of the dose-response curves [13, 14, 17, 18]. The relative combined uncertainty of the experimental dosimetry system was 6.2 % (approximately 95 % confidence levels and $k = 2$). All relative extended uncertain ties are equal because of the uncertainty of the dose detector system and standard irradiated glass dosimeters with traceability. We obtained the radiation dose using the MCNP code with a relative error, directly obtained from the code, of less than 1% [7].

Table 1 shows the dose assessment results for the laboratory mouse, 3-D mouse phantom, MCNP phantom, and MCNP simulation. A mouse apartment radia-
tion system delivered a dose of 100 mGy to the heart
Table 1. Results of the dose assessment of
the inserted glass dosimeter tom, and MCNP simulation. A mouse apartment radiation system delivered a dose of 100 mGy to the heart

Table 1. Results of the dose assessment of the inserted glass dosimeter in each target organ $\frac{1}{5}$ and

	Irradiated dose [mGy]					
Mouse model	Brain	Heart	Lung (left)	Lung (right)	Abdominal	80
Laboratory mouse	108.5	89.7	94.9	92.5	83.6	70
3-D mouse phantom	106.5	90.2	95.9	95.7	86.0	Brain
MCNP 3-D phantom	108.8	93.6	96.5	99.1	81.9	Figure 5. Com
MCNP simulation	106.0	92.5	96.9	97.5	87.9	mouse, 3-D mo MCNP simulat

RESULTS AND DISCUSSION position, and the tissue inside the mouse attenuated the from a laboratory mouse using CT scanning and $3-D$ \qquad brain, which was closer to the radiation source, rephoton. Therefore, the radiation dose that penetrated the heart was evaluated to be lower than $100 \,\mathrm{mGy}$. The ceived a radiation dose higher than 100 mGy. The abdomen, which required the radiation to penetrate deeper into the mouse, received a radiation dose of less than $100 \,\mathrm{mGy}$ owing to radiation attenuation. Each assessed dose of the target organ was within uncertainty. The most uncertainty was the uncertainty of the measuring equipment and the uncertainty of the standard irradiated glass dosimeter.

> In all dose assessment cases, the distance between the radiation source and heart position was set to 2 m , and 100 mGy was irradiated from this distance. The linear algebra-MCNP phantom shows a slightly higher dose value in the lungs, and the MCNP simulation shows a slightly higher dose value in the abdominal cavity. These differences are caused by the different geometric structures of the laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simulation.

> The dose assessment results for each mouse model are shown in fig. 5. The results of all mouse models show a measured dose value higher than 100 mGy in the brain, where the distance between the radiation source and target organ was closer than the reference position, and the measured dose value, in all mouse models, decreased toward the abdominal cavity. In the case of the heart organ, it was wrapped in the lungs, which tended to be lower than the dose evaluated in the lungs.

CONCLUSSION

This study aimed to accurately measure the radiation doses that reach target organs-a process that is essential for ensuring the reliability of pre-experiments, which are required before evaluating the biological effects of radiation. The results from the laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simu-

Figure 5. Comparison of dose assessment for laboratory mouse, 3-D mouse phantom, MCNP 3-D phantom, and MCNP simulation at center position

lation show that dose values higher than 100 mGy were measured in the brain, where the distance between the radiation source and target organ was closer than the refer-

[1] Hann, S. Y., et al., Recent Advances in 3-D Printing: ence position. The dose value decreased toward the abdomen in all cases. The uncertainties of final dosimetry include the dose detection system and dosimetry system. 46-63 The measured dose evaluation values in the target organs $[2]$ Rebby, V. S., *et al.*, Contemporary Standpoint and Fuof the laboratory mouse and 3-D mouse phantom differed by up to 3 %. Moreover, the measured dose values of a 3-D mouse phantom are within the measurement un-
 $\begin{bmatrix} 3 \end{bmatrix}$ Sreekala, P., *et al.*, 3-D Organ Printing: Review on certainty for each target organ. This shows that the laboratory mouse may be replaced with the 3-D mouse phan-
tom for dose evaluation of target organs in advance or in [4] Tian, F., et al., Radiopharmaceutical Imaging Based tom for dose evaluation of target organs in advance or in $\begin{bmatrix} 4 \end{bmatrix}$ real-time. However, the 3-D mouse phantom does not reflect the movement of a living laboratory mouse. In the future, the movement of living laboratory mice will be $\begin{bmatrix} 5 \end{bmatrix}$ Entezam, A., et al., Investigation of Scattered Dose in a evaluated and applied to dose assessment using a 3-D mouse phantom. Furthermore, in the future, the MCNP simulation will be directly compared with the manufac-
tured 3 D mouse phontom using the DICOM file ob [6] Antoniu, A., et al., Development of an US, MRI, and tured 3-D mouse phantom using the DICOM file obtained through CT scanning. The MCNP simulations and phantoms in this study were representative of mice, but
geometric simulations of detailed organs were not possi-
[7] Pelowitz, D. P., MCNPX User's Manual, Version geometric simulations of detailed organs were not possi- [7] ble. Therefore, the geometry of linear algebra-based phantoms needs to be improved to voxel-based phan-

[8] $\begin{bmatrix} \text{EA-CP-11-00438}, \text{Los Alamos, N. Mex, USA, 2011} \\ \text{R3} \end{bmatrix}$ $\begin{bmatrix} \text{EA-CP-11-00438}, \text{Los Alamos, N. Mex, USA, 2011} \\ \text{R4-CP-11-00438}, \text{Los Alamos, N. Mex, USA, 2011} \end{bmatrix}$ toms, and studies of MCNP representative phantoms de-
ment of Prospective Cancer Risks from Occupational velopment are also required for direct comparison with 3-D printed phantoms.

This work was supported by the Dongnam Institute of Radiological & Medical Sciences (DIRAMS) | 372-379 grants $(50491-2024)$ and grants from the National Research Foundation (NRF-2020M2C8A2069351) funded
hy the Ministry of Science and ICT (MSIT) [12] Hindorf, C., *et al.*, Evaluation of Parameters Influencby the Ministry of Science and ICT (MSIT).

The idea for this study was initiated by Y.-R. Kang, W. S. Jo, C. G. Lee, and Y. U. Kye. Experiments, [14] $*$, Korea Association of Standards & Testing Organidata collection, and statistical analysis were carried
zations, Standard Calibration Procedure of Ionization out by M. J. Bae, H. J. Jang, S. Mok, Y. U. Kye, H. J.

Kim. and J. E. Lee. Y.-R. Kang supervised presented [15] ***, International Organization for Standardization, Radia-Kim, and J. E. Lee. Y.-R. Kang supervised presented [15] re search and helped with its development that resulted in this paper. All the authors participated in the discus-
 $[16]$ No, S. J., *et al.*, Evaluation of in Vivo Low-Dose sion of the presented results.

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REFERENCES

- e....

121-126 125
 REFERENCES

[1] Hann, S. Y., *et al.*, Recent Advances in 3-D Printing:

Vascular Network for Tissue and Organ Regenera-

tion, *Translational Research*, 211 (2019), Sept. pp.

46-63

[2] Rebby, V. S. Vascular Network for Tissue and Organ Regeneration, Translational Research, 211 (2019), Sept. pp. 46-63 **REFERENCES**

121-126 125
 REFERENCES

125

125

121-126

121

12019

12019

12019

12019), Sept. pp.

146-63

22

Rebby, V. S., *et al.*, Contemporary Standpoint and Fu-

140-63

12019 Rebby, V. S., *et al.*, Contempor [3] Sreekala, P., et al., 3-D Or gan Print ing: Re view on **EMPLET ASSET 125**

125
 REFERENCES

125
 REFERENCES

121

Hann, S. Y., et al., Recent Advances in 3-D Printing:

Vascular Network for Tissue and Organ Regenera-

16-63

12] Rebby, V. S., et al., Contemporary Standpoi **EXECT:** 125

121-126

125
 REFERENCES

125

125
 REFERENCES

121

Vascular Network for Tissue and Organ Regenera-

tion, *Translational Research*, 211 (2019), Sept. pp.

14-63

121 Rebby, V. S., et al., Contemporary
- ture of 3-D Bioprinting in Tissue/Organs Printing, Current Opinion in Biomedical Engineering, 27 (2023), Sept., 100461
- Operational Challenges and Constraints, Materials Today: Proceeding, 33 (2020), 7, pp. 4703-4707
- on 3-D-CZT Compton Camera with 3-D-Printed Mouse Phantom, Physica Medica, 96 (2022), Apr. pp. 140-148
- Mouse Phantom Model for Pre-Clinical Dosimetry Studies, Radiation Physics and Chemistry, 189 (2021), Dec., 109691
- **ETALLACES**

125

125

127-126

127-1206

127-1206

127-1206

127-1207-1207

128-1207

128-63

127-1207-1207-1207-1207-1207-1207-1201

146-63

127-1207-1207-1207-1207

146-63

128-bby, V. S., et al., Contemporary Standpoi CT Imaging Compatible Realistic Mouse Phantom for Thermal Ablation and Focused Ultrasound Evaluation, Ultrasonics, 131 (2023), May, 106955 **REFERENCES**

[1] Hann, S. Y., et al., Recent Advances in 3-D Printing:

Vascular Network for Tissue and Organ Regenera-

tion. Translational Research, 211 (2019), Sept. pp.

16-63

[2] Rebby, V. S., et al., Contemporary [1] Hann, S. Y., *et al.*, Recent Advances in 3-D Printing:

Vascular Network for Tissue and Organ Regenera-

160. *Translational Research, 211* (2019), Sept. pp.

46-63

Rebby, V. S., *et al.*, Contemporary Standpoint an 140-18

Elsebby, V. S., et al., Contemporary Standpoint and Fu-

Re-63

tores of 3-D Bioprinting in Tissue/Organs Printing. Cur-

true of 3-D Bioprinting in Tissue/Organs Printing. Cur-

ter na tionic and Com mission of C and the original in the saccorigas Fundancy

rent Opmion in Biomedical Engineering, 27 (2023),

Speckala, P., et al., 3-D Organ Printing: Review on

Operational Challenges and Constraints, Mareria's

Today: Proceeding, 33 Protational Contenting Carrier (Fig. 1) For *all,* Radiopharmeterical Imaging Based
on 3-D-CZT Compton Camera swith 3-D-Printed
on 3-D-CZT Compton Camera swith 3-D-Printed
Mouse Phantom, *Physica Medica*, 96 (2022), Apr.
- 2.7.0, Los Alamos National Laboratory Report LA-CP-11-00438, Los Alamos, N. Mex, USA, 2011
- Exposure to Ionizing Radiation, IAEA-TECDOC-
- 1985, Vienna, Austria, 2021
***, International Commission on Radiological Protection, Low-Dose Extrapolation of Radiation-Related Cancer Risk, ICRP Publication 99, 2005
- ACKNOWLEDGEMENT

[10] Lee, J.-S., et al., Filament Material Evaluation for Breast Phantom Fabrication Using Three-Dimensional Printing, Nucl Technol Radiat, 35 (2020), 4, pp.
	- [11] ***, Materialise NV, 3-D Medical Image Processing Software, Materialise Mimics, Belgium, 2013
	- ing S Values in Mouse Dosimetry, The Journal of Nuclear Medicine, 45 (2004), 11, pp. 1960-1965
- AUTHORS' CONTRIBUTIONS [13] ***, International Atomic Energy Agency, Absorbed
Dose Determination in External Beam Radiotherany [12] Hindorf, C., et al., Eval u a tion of Pa ram e ters In flu enc - [5] Enterzam, A., *et al.*, Investigation of Scattered Dose in a

Mouse Phantom Model for Pre-Clinical Dosimetry

Studies, *Radiation Physics and Chemistry*, *189* (2021),
 (6) Antoniu, A., *et al.*, Development of an U Dose Determination in External Beam Radiotherapy: An International Code of Particle for Dosimetry Based on Standards of Absorbed Dose to Water, TRS-398, Vienna, Austria, 2004 [17] μ_1 , Computer (manner) and Forcester Material Absolute and Forcester Manual Version and Forcester Stan Material Absolute 2.7.0. D. S. MCNPX User's Manual Version 2.70. Los Alamos National Laboratory Report LA-CP-1 [7] $F(20, 20, 10, 20, 20)$, In term and Laboratory Report 2.7.0, I.e. Alternational Actratory Report LA-CP-11-00438, Los Alamos, N. Mex, USA, 2011 [8] ***, International Atomic Energy Agency, Assessmement of Prospective [8] $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ (a $\frac{1}{2}$ $\frac{1}{2}$ (b) $\frac{1}{2}$ (S) $\frac{1}{2}$ (S) $\frac{1}{2}$ (D) (S) $\frac{1}{2}$ (D) (S) $\frac{1}{2}$ (D) (S) $\frac{1}{2}$ Exposite to interactional Commission on Radiological Prosession
1985, Vienna, Austria, 2021

[9] ***, International Commission on Radiolano-Re-

lead Cancer Risk, ICRP Publication 99, 2005

Lee, J.-S., *et al.*, Filament
	- Chamber Dose Meter, KASTO 20-80105-042, 2020
	- tion Protection Performance Criteria for Radiobioassay, ISO 28218, 2010
	- Mouse Irradiation System, Journal of Instrumenta-
	- tion, 11 (2016), P03031
[17] ***, International Organization for Standardization/ /International Electrotechnical Commission, Uncertainty of Measurement-Part3: Guide to Expression of Uncertainty in Measurement, ISO/IEC GUIDE 98-3, 2008
	- Errical Primary Based Districts (Englishmonton Comparison)

	First Phanon Fabricaliso Nivi 3, 1923-379

	[11] ***, Materialise Nimites, Belgium, 2013

	[12] Hindorf, C., etal., Evaluation of Parameters Influence

	[12] Hindor tion/American Society for Testing and Materials, Practice for Calibration of Routine Dosimetry Systems for Radiation Processing, ISO/ASTM 51261: 2013(E), 2013

Re ceived on July 30, 2024 Accepted on October 9, 2024

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PA3BOJ 3-D IIITAMIIAHOF ФАНТОМА МИША КАО ЗАМЕНЕ ЗА САДАШЊИ ЖИВОТИЊСКИ МОДЕЛ МИША

Процена дозе зрачења циљаних органа лабораторијског миша захтева да се стаклени дозиметар хируршки убаци на озрачено место. Међутим, прецизно уметање стакленог дозиметра на исту локацију код различитих мишева ретко се постиже, смањујући поузданост измерене дозе зрачења. Да би се решило ово ограничење, развијен је 3-D фантом миша коришћењем СТ скенирања и технологије 3- D штампања. Доза зрачења циљаних органа процењена је коришћењем четири модела миша: лабораторијског миша, 3- Dфантома миша, MCNP 3- Dфантома и MCNP симулације. У свим експериментима, мозак, срце, плућа и абдомен зрачени су са 100 mGy измерене керме у ваздуху при јачини керме од 6 mGyh⁻¹. Стаклени дозиметар мале запремине убачен је у моделе миша да би се проценила доза зрачења, а поузданост система за очитавање стакленог дозиметра процењена је коришћењем криве доза-одзив. Утврђено је да се вредности дозе лабораторијског миша и 3-D штампаног фантома миша разликују до 3.3 %. Овај рад обезбеђује методу за прецизно мерење дозе зрачења на циљане органе, повећавајући поузданост припреме експеримената.

 K ључне речи: сійандардно озрачивање, дозимешрија са сійаклом, 3-D фанійом миша, M он \bar{u} е Карло симулација